

# **Request for input from the ISCEV membership: ISCEV standard EOG (2017) Update -- October 2016 Draft**

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Overall there has been no change to the testing protocol of the Standard EOG, 15 minutes dark then 15 minutes light with testing of 10 saccades each minute. The Fast Oscillation is a recommended test that uses continuous recording with 60 second intervals of light and dark for a minimum of four light-dark cycles.

## **Summary of Major Changes**

1. Updated and extended sections covering the mechanism of the EOG and FO are given in this new draft based on the developments in our understanding of bestrophin and as a background to the different mechanism of these electrophysiological tests of RPE function.
2. The new draft allows of stopping the EOG once a light-rise has been obtained so that the standard EOG ratio can be calculated, in an effort to minimize the length of the procedure.
3. The new draft suggests a method for smoothing the physiological response using a weighted average of the standing potential amplitudes –although other methods for smoothing the EOG plot are acceptable.
4. A move away from the Arden Eponym, with the ‘Arden ratio’ noted but now referred to as ‘the standard EOG ratio’
5. Figures have been updated:
  - a. Figure 1 Replaced with real data showing 0.1 Hz high pass filter
  - b. Figure 2- New figure showing blink artefact and over/undershoot with correct positioning of cursers.
  - c. Figure 3- Unchanged image of the electrode positions but modified caption.
  - d. Figure 4- Raw and smoothed trace of the standard EOG indicating the Dark Trough and Light Peak
  - e. Figure 5- Trace of the Fast Oscillation.: Replaces figure 4 schematic of the FO in the 2011 Standard.
6. Reporting of the Dark Trough amplitude is recommended as before but an explanation is given now that this is an indication of the degree of the strength of the tight-junctions between the RPE cells that is proportional to the epithelial potential that is recorded as the ‘standing potential’ with the EOG.

## **Summary of Minor Changes**

- 1) Electrode placement and preparation are consistent with the VEP standard
- 2) a.c and d.c changed to AC and DC – this is the convention
- 3) Fast Oscillations are singular as this is a single oscillation

## Areas for Comments/Discussion:

### 1. Pre-Adaptation:

- a. The committee discussed the addition of a suggestion that there be a period of 2-5 minutes under 30 cd/m<sup>2</sup> ahead of the dark phase to standardize the adaptive state of the retina.
  - a. *Do any labs have evidence that this may assist with the development of the dark trough or improve reliability of the recordings?*

### 2. Recording of Saccades:

- a. The committee was inclined to remove the section that prohibited an auditory cue ahead of each saccade as we were unsure of the impact of allowing auditory cues during the recordings.
  - i. Line 255: "However, there should not be auditory cues before every alternation of the fixation lights, as this may prompt erroneous saccades before the fixation lights change."
    - a. *Should the above sentence be retained, removed or modified?*

### 3. Impedance and filtering:

- a. The committee discussed the possibility of increasing the low pass filter to 100Hz as the 30 Hz low pass may mask poor recording?
- b. The committee also discussed whether the impedance could be < 10 kΩ with up to 3-4 kΩ between electrodes given the 0.1-30 Hz will reduce any line interference.
- c. *Ultimately the committee left the recording parameters unchanged although we would welcome comments on these points from the membership.*

### 4. Normative Data:

- a. Despite several recent studies reporting normative values for the EOG and FO, the committee recommends lab based normative data be used.
  - b. *Given the standardisation of the EOG with respect to duration, electrode use and illumination the committee suggests that a normative database could be established for the EOG with reference ranges available in a future standard?*
  - c. *This may be possible through a meta-analysis of published data or across labs that would be willing to share data sets?*
-

1 **Consultation Draft October 2016**

2 **ISCEV Standard for Clinical Electro-oculography (2017**  
3 **update)**

4

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7

8 for the International Society for Clinical Electrophysiology of Vision, [www.iscev.org](http://www.iscev.org)

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10 Anthony G. Robson (Director of Standards), M Bach (Director of Communications)

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12 *For editorial use:*

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14 Address for Correspondence:

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16 This update was approved by ISCEV. This document is available on the ISCEV  
17 website: <http://www.iscev.org>.

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20 The authors represent the International Society for Clinical Electrophysiology of  
21 Vision.

22

23 **Abstract**

24

25 The clinical electro-oculogram (EOG) is an electrophysiological test of the outer  
26 retina and retinal pigment epithelium (RPE) in which changes in the electrical  
27 potential across the RPE are recorded during successive periods of dark and light  
28 adaptation. This document presents the 2017 EOG Standard from the International  
29 Society for Clinical Electrophysiology of Vision (ISCEV: [www.iscev.org](http://www.iscev.org)). This  
30 standard has been reorganized and updated, but without substantive changes to the  
31 testing protocol from the previous version published in 2011. It describes methods for  
32 recording the EOG in clinical applications and gives detailed guidance on technical  
33 requirements, practical issues, and reporting of results. It is intended to promote  
34 consistent quality of testing and reporting within and between clinical centers.

35

36 **Key words** Arden ratio, ISCEV standards, Clinical Electrophysiology, Electro-  
37 oculogram (EOG), Light adaptation, Retinal pigment epithelium (RPE), Fast  
38 Oscillation (FO)

39

40 **Abbreviations**

41 DT – Dark Trough

42 EOG – Electro-oculogram

43 ER – Endoplasmic Reticulum

44 ERG – Electroretinogram

45 FO – Fast Oscillation

46 ISCEV – International Society for Clinical Electrophysiology of Vision

47 LP – Light Peak

48 RPE – Retinal Pigment Epithelium

49 TEP – Trans Epithelial Potential

50

51

52 This Standard is one of a series of ISCEV Standards and Guidelines for  
53 clinical electrophysiology of vision [1-7] developed by ISCEV and listed here:  
54 [iscev.org/standards](http://iscev.org/standards) [8]. This Standard supersedes previous versions of the ISCEV  
55 Standard for Clinical Electro-oculography which was first published in 1993 [1] and  
56 subsequently revised and last published in 2011 [2]. This Standard contains updated  
57 information and has been reorganized for greater clarity and for consistency with  
58 other ISCEV Standards, without substantive changes to the 2011 testing protocol.

59 This ISCEV Standard describes basic procedures that allow reproducible  
60 recordings that are comparable across laboratories. It is intended that the ISCEV  
61 Standard EOG protocol be used widely, but not to the exclusion of other tests or  
62 protocols that are not covered by this Standard. Electrophysiologists are encouraged  
63 to extend the EOG as required when clinically relevant to maximize the diagnostic  
64 value of the standard EOG and FO recordings for their patients and for clinical trials.

65 Clinical and research users of the EOG are encouraged to use the most recent  
66 Standard, to achieve consistency of results within and between test centers. Reports of  
67 standard EOG recordings performed in this manner should cite this 2017 Standard.  
68 When a method is used which deviates from the Standard, the deviations should be  
69 fully described.

70

## 71 **The Electro-oculogram**

### 72 **Electrophysiology of the outer retina in dark and light**

73 There is a difference in electrical potential between the anterior and posterior  
74 of the eye, known as the standing potential of the eye. The standing potential is an  
75 indirect measure of the trans-epithelial potential (TEP) of the retinal pigment  
76 epithelium (RPE). The TEP is equal to the difference in the membrane potential of the  
77 basolateral and the apical membranes, which are electrically coupled through the  
78 tight-junctions of the RPE. Changes in the resistance between the apical or basolateral  
79 membranes or membrane potentials alter the amplitude of the TEP and thus the  
80 recorded standing potential of the eye.

81 A change in the standing potential can be induced by retinal adaptation to  
82 ambient illumination by alternating intervals of dark and light. The standard EOG  
83 utilizes the RPE's response to changing illumination to assess the function of the  
84 outer-retina and RPE. There are two clinical tests of the standing potential of the eye.  
85 The standard EOG is recorded during 15 minutes of dark adaptation followed by 15  
86 minutes of light adaptation. The Fast Oscillation (FO) is a different additional EOG  
87 procedure which is performed during alternate 1 minute dark and light periods. The  
88 changes in the standing potential for the standard EOG and the FO are derived from  
89 different mechanisms.

90

#### 91 **Mechanism of the standard EOG**

92 During the 15 minute period of dark adaptation there is a fall in the recorded  
93 standing potential typically reaching a minimum at 10–15 minutes, and this is referred  
94 to as the Dark Trough (DT). The DT amplitude is stable and thus is an indirect  
95 measure of the integrity of the RPE based on the TEP, which is maintained by the  
96 tight junctional proteins between the apical and basal membranes. For this reason the  
97 amplitude of the DT should be noted.

98 Following light onset there is an increase in the TEP of the RPE producing the  
99 light-rise of the EOG. The mechanism initiating the light-rise is unknown but it  
100 requires a normally functioning interface between the rod photoreceptors and RPE.  
101 The light-rise is ultimately the result of an increase in intracellular free calcium,  
102 which is released from the endoplasmic reticulum (ER), regulated by an interaction  
103 between ER bestrophin and L-type calcium channels associated with the basolateral  
104 membrane. Intracellular calcium gates the opening of a basolateral calcium-dependent  
105 chloride channel. Increased chloride conductance depolarizes the basolateral  
106 membrane which increases the TEP, recorded as an increase in the standing potential  
107 of the eye. The light-rise usually reaches a maximum at 7–12 minutes after light onset  
108 and is known as the Light Peak (LP). The LP is the first of several peaks (the damped  
109 oscillation) which become progressively smaller for up to 90 minutes during light  
110 exposure.

111 The standard clinical EOG provides an indirect measure of the minimum  
112 amplitude of the standing potential in the dark (at the DT), and then again at its  
113 maximum amplitude in the light (at the LP). This is expressed as the standard EOG  
114 light peak to dark trough (LP : DT) ratio, also known as the “Arden ratio”.

115

### 116 **Mechanism of the Fast Oscillation**

117 The FO is an optional additional test that has a different mechanism to the  
118 standard EOG owing to the shorter dark and light intervals used. At light onset there  
119 is a fall in potassium in the sub-retinal space that causes a strong outward potassium  
120 current across the apical membrane of the RPE and subsequent hyperpolarization, and  
121 c-wave, of the ERG. The fall in sub-retinal potassium also slows the apical NaK2Cl  
122 co-transporter and reduces intracellular levels of chloride which in turn hyperpolarizes  
123 the basal membrane and reduces the TEP and creates the trough of the FO after 35–45  
124 seconds after light onset. The TEP returns to normal, as ionic homeostasis is restored  
125 and a peak is recorded during the subsequent dark period after in a further 35–45  
126 seconds. The alternation between dark and light at one minute intervals establishes a  
127 continuous oscillation that is dependent on changes in ionic permeability at the apical  
128 and basal membranes and the electrical coupling between these membranes by tight  
129 junctions.

130

### 131 **Diseases affecting the light response of the EOG**

132 The light response of the EOG is affected in diffuse disorders of the RPE and  
133 disorders of the photoreceptor layer of the retina including acquired retinopathies and  
134 retinal dystrophies characterized by rod dysfunction or chorio-retinal atrophy. In most  
135 of these disorders EOG abnormalities are proportional to the severity of rod-mediated  
136 electroretinogram (ERG) abnormalities. Notable exceptions include disorders of the  
137 bestrophin gene (*BEST1*). These include Best vitelliform macular dystrophy (Best  
138 disease), autosomal recessive bestrophinopathy (ARB), and autosomal dominant  
139 vitreoretinopathopathy (ADVIRC). In Best disease the ERG is usually normal and  
140 the standard EOG ratio abnormal. A normal EOG may distinguish Best disease from

141 other autosomal dominant retinal disorders with similar fundus features including  
142 some cases of adult vitelliform macular dystrophy and pattern dystrophy. In ARB and  
143 ADVIRC the ERG is often abnormal but the EOG is severely or disproportionately  
144 abnormal. An abnormal EOG, not explained by ERG reduction, may also be  
145 associated with some toxic and other retinopathies.

146

### 147 **Principles of the clinical EOG measurement**

148 The standing potential of the eye may be assessed using skin electrodes  
149 attached either side of each eye to record successive horizontal saccadic eye  
150 movements. The patient tracks alternating lights separated by a fixed angle, to enable  
151 constant eye movement excursions which are recorded as a series of positive and  
152 negative deflections or potentials that coincide with ocular rotation. The magnitude of  
153 the eye movement potential (at a fixed angle) is a fixed proportion of the standing  
154 potential. During a dark/light cycle, the magnitude of the potential changes  
155 proportionate to changes in the TEP across the RPE.

156

### 157 **The Standard Method**

#### 158 **Technologic requirements**

##### 159 **Electrodes**

160 Skin electrodes such as sintered silver–silver chloride, standard silver–silver  
161 chloride or gold cup electrodes are recommended for recording EOGs. The skin  
162 should be prepared by cleaning, and a suitable paste or gel used to ensure good, stable  
163 electrical connection. The electrode-skin contact impedances should be below 5 k $\Omega$  as  
164 measured between 20 and 40 Hz.

165

##### 166 **Stimulator**

167 The EOG should be performed using a full-field (Ganzfeld) dome. The  
168 Ganzfeld should have a comfortable head/chin rest, and two red fixation lights located

169 15° left and right of center. The fixation lights should be bright when the light  
170 adapting background is on, and dim (to be just visible) in the dark.

171

### 172 **Light and dark**

173 The dark phase should take place in total darkness, with the fixation lights  
174 dimmed to the minimum necessary to enable fixation.

175 The light phase requires even lighting throughout the dome. The adapting light  
176 should appear white and have a luminance of 100 photopic  $\text{cd/m}^2$  measured at the  
177 position of the eye. To account for minor variability in equipment and calibration the  
178 acceptable range within the Standard for the light adapting background is 90 to 110  
179 photopic  $\text{cd/m}^2$ . Calibration of the Ganzfeld stimulator should be carried out  
180 periodically. Modest room lighting may be turned on during the light phase as long as  
181 ambient luminance is less than that in the bowl.

182 Note that adapting light sources of different types, such as tungsten, halogen,  
183 LED, and fluorescent, have different spectral characteristics and the color may change  
184 with brightness. This makes the definition of standard lighting inherently imprecise,  
185 although for practical purposes most “white” light of the correct luminance will give  
186 very similar results.

187

### 188 **Amplification and filtration**

189 Amplifiers should have a band pass of either 0 to 30 Hz (DC), or 0.1 to 30 Hz  
190 (AC), to make recordings of saccadic eye movements appear as square waves (figure  
191 1). For a 30° saccade, the typical standard EOG amplitude is between 250 and 1000  
192  $\mu\text{V}$  with an essential frequency content of 0 to 30 Hz. In theory, the ideal recording  
193 technique is DC amplification but this is generally impractical because of baseline  
194 drift. Thus, we recommend AC recording with a 0.1 Hz high pass filter. If a higher  
195 frequency filter is used (e.g. 0.5 Hz), it will distort the square waves, making  
196 identification of overshoot and stepped saccades difficult.

197

198 \_\_\_\_\_ Insert Figure 1 Near Here \_\_\_\_\_

199

200 Manufacturers should allow the examiner to have access to all of the raw data  
201 for each 10 second recording epoch so that the examiner is able to visually inspect the  
202 individual saccadic records to enable accurate cursor placement around any artefacts  
203 to measure the standing potential as shown in figure 2.

204

205 \_\_\_\_\_ Insert Figure 2 Near Here \_\_\_\_\_

206

207 The operator should be able to see the saccadic recordings as they occur, to  
208 ensure that there are no artefactual signals, problems such as amplifier saturation (that  
209 can be corrected by adjusting amplifier gain), and that patients are compliant with the  
210 task.

211

## 212 **Preparation of the patient**

### 213 **Pupils**

214 The pupils should be dilated maximally before testing and their size recorded.  
215 If full pupil dilation is impossible or undesirable, an attempt should be made to  
216 increase the adapting luminance so that an equivalent retinal illumination is  
217 approximated. The amount of light passing through the pupil, when measured in  
218 Trolands, is the product of luminance (in  $\text{cd/m}^2$ ) and pupil area (in  $\text{mm}^2$ ). For  
219 example, to produce the same effect upon the retina, twice as much dome luminance  
220 is required with a 5-mm diameter pupil (roughly  $20 \text{ mm}^2$ ) than with a 7-mm diameter  
221 pupil (roughly  $40 \text{ mm}^2$ ). The report should describe any deviations from the Standard.

222

### 223 **Electrode placement**

224 After suitable skin preparation, recording electrodes should be placed close to  
225 the canthi of each eye as in Figure 3. The electrodes from each eye are connected to

226 separate channels of a differential amplifier. A separate electrode should be attached  
227 and connected to the ground. Convenient and commonly used ground electrode  
228 positions include the forehead, vertex, mastoid and earlobe. The impedance between  
229 any pair of electrodes should not exceed 5 k $\Omega$ . The electrodes, amplifier and  
230 impedance meter must be approved for medical use.

231

232 \_\_\_\_\_Insert Figure 3 Near Here\_\_\_\_\_

233

### 234 **Pre-exposure to light**

235 The patient should be in stable indoor lighting before the test, to stabilize the  
236 EOG, for at least 30 minutes. Indirect ophthalmoscopy, fundus autofluorescence  
237 and/or photography and other strong illumination changes must be avoided during this  
238 period. As near as practical, the pre-test light exposure should be the same for all  
239 patients.

240

### 241 **Explanations**

242 The procedure should be explained to the patient, noting that head position  
243 must not change, as this is one common artefact source, and that the eyes should only  
244 move left and right. The patient should be instructed not to anticipate the onset of the  
245 alternation of the fixation lights but to move their eyes only when the lights change.  
246 Practice the procedure with the recording system prior to dark adaptation, to  
247 familiarize the patient with the task and to check on the stability and quality of the  
248 recorded EOG.

249

### 250 **Clinical recording**

#### 251 **Recording of saccades**

252 Fixation lights should alternate once per second, for 10 seconds out of every  
253 minute. The EOG potentials are recorded during these 10 second periods. There

254 should be a warning, verbal or automatic, of the impending start of each recording  
255 period to ensure readiness of the patient and operator. However, there should not be  
256 auditory cues before every alternation of the fixation lights, as this may prompt  
257 erroneous saccades before the fixation lights change.

258

### 259 **Dark phase**

260 For the dark phase total darkness should be maintained for 15 minutes, except  
261 for the dim fixation lights. EOG recordings should be made once a minute for 10  
262 seconds, as specified above. The operator should have a concurrent view of the  
263 recordings to check for patient compliance, and for errors such as noise or overshoot.

264

### 265 **Light phase**

266 For the light phase a dome background light of 100 photopic  $\text{cd/m}^2$  should be  
267 turned on to “trigger” the initial light phase response and should remain on for the  
268 duration of the light phase. However, the luminance can be increased gradually over a  
269 short period (e.g. 20 seconds) to minimize patient discomfort. Longer ramps (e.g.  
270 lasting minutes), will alter the responses. Continue the recording for 10 seconds out of  
271 every minute (as above) for at least 15 minutes to register the presence or absence of  
272 the LP. If the LP can be clearly identified during the recordings then the test may be  
273 stopped before 15 minutes. It may be necessary to extend the light phase to fully  
274 characterize abnormal responses but a delay in the LP should be reported. The patient  
275 should remain positioned in the headrest of the stimulator bowl throughout the  
276 procedure, with eyes open to maintain retinal illumination.

277

### 278 **Patient compliance**

279 Patients will have difficulty performing saccadic eye movements if they  
280 cannot fixate reliably because of poor central vision, diplopia or ocular motility  
281 problems (including nystagmus). Patients with diplopia may be advised to look  
282 between the pair of images, or one eye can be patched if the suspected retinal disorder

283 is bilateral. Patients who are very young or those with a physical or learning disability  
284 may not be able to perform the standard EOG. In young children with suspected Best  
285 disease it may be possible to test their parents, since a carrier of Best disease will have  
286 an abnormal EOG light-rise, irrespective of whether the fundus is normal.

287 Patient compliance can vary due to fatigue or inattention. Common problems  
288 are drifting of the head back from the stimulator, head turning, and irregular eye  
289 movements during the recording or eye closure during the light phase. These can be  
290 minimized by having a real-time view of the EOG recording via an infrared camera.  
291 In most cases, gentle coaching and reminders will minimize the effects of poor  
292 compliance.

293

## 294 **Analysis and reporting**

### 295 **Saccadic amplitude**

296 The EOG amplitudes should be measured in microvolts ( $\mu\text{V}$ ) either manually  
297 or by a computer algorithm after visual inspection. Care must be taken to avoid  
298 measuring the effects of overshoot (see Figure 2) or irregular (artefactual) saccades.  
299 The average of the amplitudes within each 10-second recording epoch should be  
300 measured, excluding outliers, artifacts or responses consistent with poor fixation. If a  
301 computer algorithm is used, it is important to ensure that the values returned represent  
302 true EOG amplitudes and not artefactual records. Particular causes of unreliability are  
303 overshoot, stepped saccades, missing saccades, inverse saccades (eyes going in the  
304 opposite way to the fixation lights), and eccentric fixation in which the saccade length  
305 switches between two or more values.

306

### 307 **Dark trough and light peak: Smoothing**

308 The average EOG amplitude calculated from each 10 second epoch should be  
309 plotted. However, there is always “noise” in real-life recordings, and the goal of the  
310 EOG measurement is to record the physiological DT and LP, rather than the lowest or  
311 highest values. Thus, the first critical step is that the underlying physiologic curve be

312 recognized and drawn, in order to derive the DT and LP amplitudes. This can be  
 313 achieved by using a curve fitting algorithm, a spline (curve fitting ruler) or the  
 314 weighted average of each point where the Amplitude ( $A$ ) at time point ( $t$ ) is given by  
 315 equation 1;

316

$$317 \quad A_t = \frac{(A_t + (0.5A_{(t-1)}) + (0.5A_{(t+1)}))}{2} \quad \text{equation 1}$$

318

319 Figure 4 shows raw data plotted and the subsequent smoothing of the data  
 320 using equation 1. It is helpful if uncertain or artefactual values can be identified and  
 321 marked at the time of recording, so that they can be ignored later when curve-fitting.

322

323 \_\_\_\_\_Insert Figure 4 Near Here\_\_\_\_\_

324

### 325 **Light peak to dark trough ratio calculation**

326 The ratio of the LP : DT amplitudes is the standard EOG ratio and also known  
 327 as the “Arden ratio”. The LP and DT amplitudes are derived from the smoothed  
 328 (physiologic) EOG record. It is important to note that a normal ratio does not imply a  
 329 normal DT and that the DT amplitude should also be reported.

330

### 331 **Reporting**

332 Clinical reports should state the standard EOG ratio, DT amplitude (in  
 333 microvolts), the time from the start of the light phase to the LP (if present), the pupil  
 334 size, and the type of adapting light source. The report should also describe any  
 335 difficulties encountered during testing that may affect confidence in the results, such  
 336 as patient compliance or inconsistent eye movements.

337

### 338 **Normative data**

339 Each center must establish its own set of normative values for the EOG and  
340 FO. The median value (not the mean) should be used to define reference values and  
341 the actual values on either side of the median that bracket 90 percent of the reference  
342 ranges (5<sup>th</sup> – 95<sup>th</sup> centile) should be determined by direct tabulation of the reported  
343 results. The normal standard EOG ratio is typically between 1.7 and 4.3 with a LP  
344 time ranging from 7-12 minutes.

345

#### 346 **Interaction between eyes**

347 The EOG potentials from one eye can contaminate the response from the  
348 other. The magnitude of this effect is approximately 15% with electrodes placed on  
349 each side of the nose close to the inner canthi. It rises to about 40% if the electrodes  
350 come close together and touch (i.e. become a common central electrode on the bridge  
351 of the nose). This interaction can give misleading results in cases of an electrically  
352 inactive eye (e.g. total retinal detachment), or absent eye, because the defective eye  
353 will appear to have the same standard EOG ratio as the fellow eye, albeit with a much  
354 smaller standing potential. If the eyes have similar standing potentials but different  
355 standard EOG ratios the measured ratio from the better eye is enhanced at the expense  
356 of that from the weaker eye, which can in fact appear to have a standard EOG ratio of  
357 less than 1.0, solely due to interaction.

358

#### 359 **Deviation from the Standard**

360 This Standard represents a basic and core procedure for the assessment of  
361 generalized function of the RPE/photoreceptor interface. If a center chooses  
362 techniques which vary from the Standard, it is critical to cite this document and  
363 specify any deviations, such as differences in the luminance level for the adapting  
364 light or in the duration of the dark or light adapting intervals. The standing potential  
365 can also be used to monitor eye-movements in studies unrelated to retinal and RPE  
366 pathophysiology.

367

### 368 **Fast Oscillation**

369           Some centers measure the fast oscillation (FO), in conjunction with the  
370 standard EOG. The FO has the opposite polarity to the standard EOG. Light onset  
371 causes a decrease in the standing potential, while in darkness there is an increase in  
372 the standing potential.

373           The FO is recorded using the same technical specifications as the standard  
374 EOG (amplifier, electrode placement, fixation targets, background luminance and 1  
375 per second saccades). However, the saccades and the recording should be continuous  
376 without interruption for the duration of the test. Light and dark are alternated every 60  
377 seconds to induce the FO, which have a near sinusoidal appearance (figure 5). During  
378 each light interval a trough develops and begins to rise again after 35-45 seconds. The  
379 subsequent interval of darkness results in a peak at 35-45 seconds following the onset  
380 of darkness. The total number of light-dark intervals should be at least 4 with 60  
381 second intervals of light and dark, making a total test time of 8 minutes. Pre-  
382 adaptation does not affect the FO, so this test can be performed either independently  
383 or in conjunction with the standard EOG.

384           Figure 5 shows a FO recording formed by dark and light intervals. The FO  
385 amplitude can be calculated from the average of the peak to trough ratios from each of  
386 the dark/light cycles. The time to the peak or trough should be calculated from the  
387 time of light offset or onset respectively and averaged for the number of light/dark  
388 cycles in the recording.

389

### 390 **Acknowledgements**

391

392

### 393 **References**

- 394 1. Marmor MF, Zrenner E (1993) Standard for clinical electro-oculography.  
395 International Society of Clinical Electrophysiology of Vision. Doc Ophthalmol  
396 85(2):115-124

397

398 2. Marmor MF, Brigell MG, McCulloch DL, Westall CA, Bach M (2011) ISCEV  
399 Standard for Clinical Electro-oculography (2010 Update). Doc Ophthalmol  
400 122(1):1-7

401

402 3. Brigell M, Bach M, Barber C, Moskowitz A, Robson J (2003) Guidelines for  
403 calibration of stimulus and recording parameters used in clinical  
404 electrophysiology of vision. Doc Ophthalmol 107(2):185-193

405

406 4. McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R,  
407 Bach M (2015) ISCEV Standard for full-field clinical electroretinography (2015  
408 update). Doc Ophthalmol 130(1):1-12

409

410 5. Hood DC, Bach M, Brigell M, Keating D, Kondo M, Lyons JS, Marmor MF,  
411 McCulloch DL, Palmowski-Wolfe AM (2012) ISCEV Standard for clinical  
412 multifocal electroretinography (2011 edition). Doc Ophthalmol 124(1):1-13

413

414 6. Bach M, Brigell MG, Hawlina M, Holder GE, Johnson MA, McCulloch DL,  
415 Meigen T, Viswanathan S (2013) ISCEV standard for clinical pattern  
416 electroretinography (PERG) – 2012 update. Doc Ophthalmol 126(1):1-7

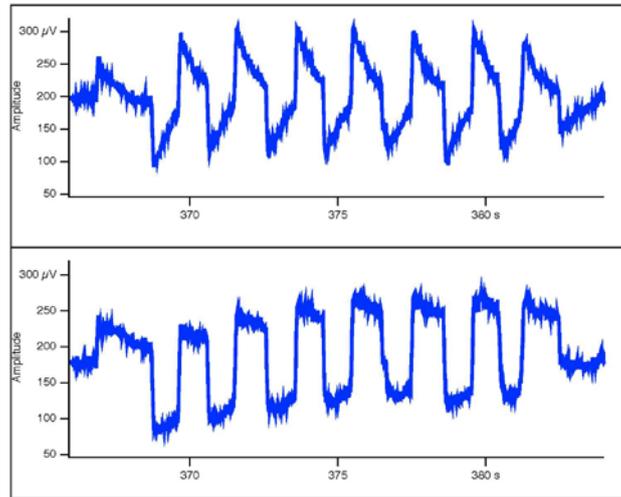
417

418 7. Odom JV, Bach M, Brigell M, Holder GE, McCulloch DLL, Mizota A, Tormene  
419 AP (2016) ISCEV standard for clinical visual evoked potentials – (2016 update).  
420 Doc Ophthalmol 133(1):1-9

421

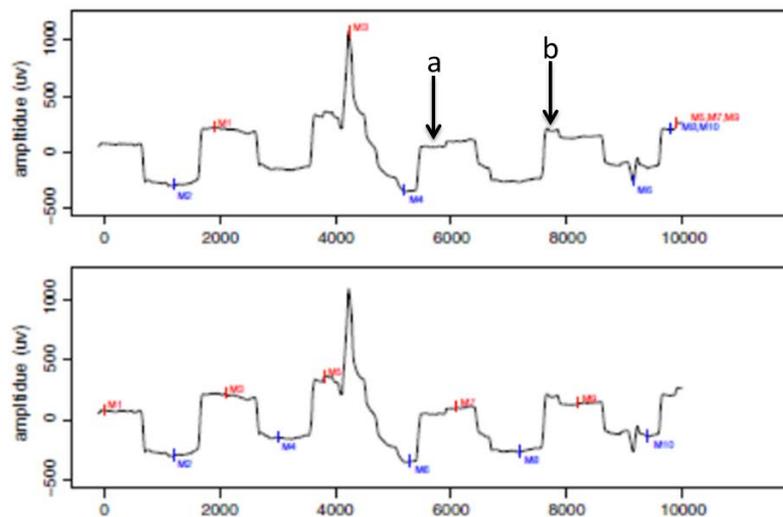
422 8. ISCEV Standards (26/10/2014). Retrieved 25/10/2016, from  
423 <http://www.iscev.org/standards/>.

424

425 **Figures and Captions**

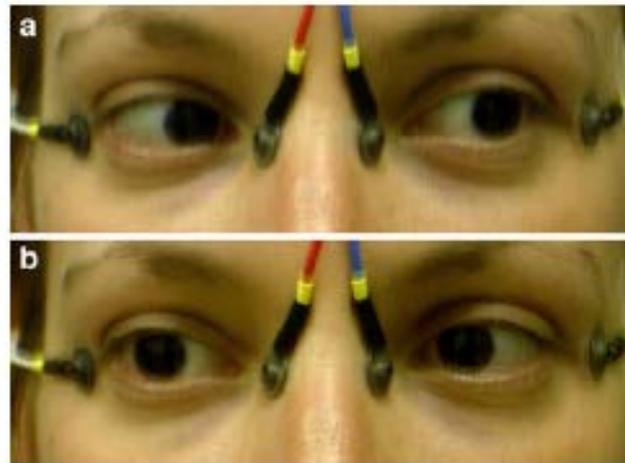
426

427 **Figure 1** Upper trace shows filtering with 0.1–30 Hz, lower trace with post-hoc DC  
 428 restoration by digital integration, rendering it similar to direct DC recording. DC  
 429 recording (or restoration) makes it easier to perform the plateau measurements.



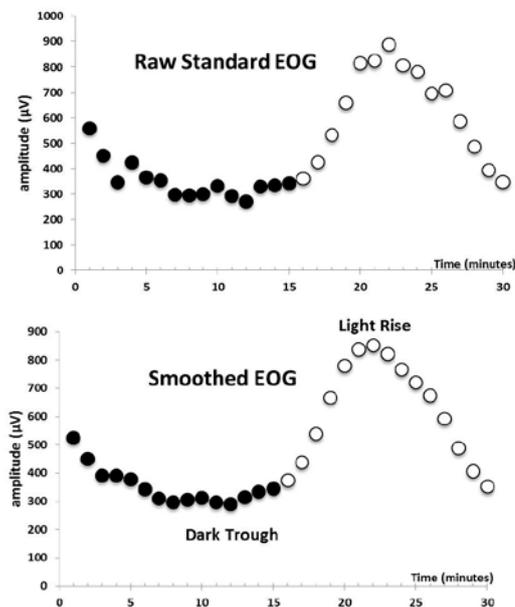
430

431 **Figure 2** Examples of 10-second saccadic records with a blink artefact at  
 432 approximately 4000 ms. Arrow ‘a’ indicates an initial undershoot and arrow ‘b’ an  
 433 overshoot of the fixation target visible by the step in the plateau of the EOG recording  
 434 (upper trace). The lower trace shows the manual placing of markers at the peak and  
 435 trough of the EOG recording as the eye performs horizontal saccades for ten seconds  
 436 at one second intervals.



437

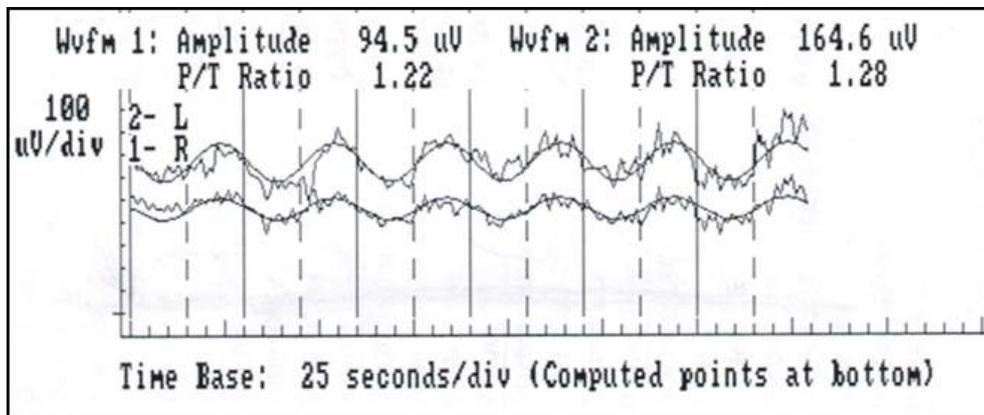
438 **Figure 3** Recording electrode positions located near the inner and outer canthi of each  
 439 eye. As the eyes perform horizontal saccades: left (a) then right (b) the amplitude of  
 440 the standing potential is recorded across the active electrodes.



441

442 **Figure 4** Upper figure shows the raw standing potential values for an EOG with 15  
 443 minutes of dark (black circles) and 15 minutes of light (white circles). Smoothing of  
 444 the data using equation 1 helps to define the physiological response and the DT and  
 445 LP points from which to calculate the standard EOG light rise to dark trough ratio  
 446 (Lower figure). Computer algorithms or fitting with a spline rule may also be utilized.

447



448

449 **Figure 5.** Typical Fast Oscillation recording with six cycles of 75 second light and 75  
450 second dark intervals generating a trough in the light and a peak in the dark interval  
451 for the Left (upper) and Right (lower) waveforms. Parameters were derived from the  
452 fit of the sine wave to the raw data. Amplitude was calculated from the difference  
453 between the absolute amplitudes of the peak and trough of the sine wave and the P/T  
454 (Fast Oscillation ratio) is the ratio of the peak to trough amplitude.