### 1. The long duration flash ERG protocol.

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## 2. Scope and applications:

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This guideline describes an extended full-field clinical electroretinography (ERG) protocol for the recording of the long-duration flash ERG (On-Off ERG). The basic technique is already widely used as an aid to diagnosis of various retinal disorders and may be recorded on most modern commercially available equipment, usually as an addition to the ISCEV-standard ERG protocol [1].

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The standard light-adapted (LA) 3.0 ERG b-wave is evoked with flashes of a short duration (<5 ms) on a rod-saturating background, and largely reflects overlapping contributions from the On- and Off- bipolar cell pathways. Separation of the function of the On- and Offpathways requires long-duration stimuli (e.g. 150-200 ms) in the presence of a rodsaturating background. The long-duration ERG has two major components, the On-response and the Off- response. The On- response occurs after the stimulus onset, and consists of two prominent waves, the negative polarity a-wave and the positive b-wave. The Off- response, or d-wave, is a positive polarity component in response to stimulus offset [2–4]. The sources of On- and Off- responses were elucidated by experimental pharmacological studies in nonhuman primates, whose ERGs are very similar to those in humans. These studies showed that the a-wave of the On-response originates from cone photoreceptors, with a significant contribution from Off- (hyperpolarizing) bipolar cells [5]. The b-wave of the On- response reflects the function of the On- (depolarizing) bipolar cells, although its amplitude and shape may be influenced by Off- bipolar and horizontal cells [6]. The d-wave is a complex response; the initial rapid phase originates from Off- bipolar cell activity, but cone photoreceptors contribute to the later slow phase and On- bipolar cells act in an opposite direction [7, 8].

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Common diagnoses that may benefit from additional On-Off ERG testing include retinal dystrophies and retinal disorders that cause dysfunction post-phototransduction or at a post-receptoral level. The On-Off ERG allows evaluation of the relative or selective involvement of On- and Off- pathways, not fully enabled by the standard LA 3.0 ERG responses [2-4, 9-15]. Common forms of congenital stationary night blindness (CSNB) are good illustrative examples. In complete CSNB there is generalised On- bipolar cell dysfunction and the waveform shows an electronegative On- response but a preserved Off-response. In contrast, incomplete CSNB is associated with abnormalities affecting both the On- and Off- responses [2]. Other retinal disorders associated with selective On- pathway dysfunction include melanoma associated retinopathy, early cases of phosphomannomutase deficiency (PMM2-CDG) [12], and some forms of autoimmune retinopathy [15]. Long duration ERGs may also be useful in X-linked retinoschisis, Batten disease, Duchenne muscular dystrophy, spinocerebellar degeneration, quinine toxicity and other disorders [16, 17].

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#### 3. Identification:

- 45 Protocol number: 1; v3 061117
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# 4. Patient population for whom the protocol is targeted:

Patients of all ages able to tolerate Ganzfeld stimulation, referred for investigation of possible retinal dysfunction, especially those with decreased light-adapted and/or dark-adapted ERG b-wave and relatively preserved a-wave (electronegative ERG or low b:a ratio), suggesting dysfunction post-phototransduction or at the level of the inner retina.

#### 5. Technical issues:

The long-duration ERG protocol will follow the specifications of the current ISCEV standard ERG [1]. Additional considerations include the following:

a) Stimulus duration. The duration of the light stimulus should be long enough to separate the On- and Off- responses. Most studies used durations of between 150-200 ms (Table 1), based on the largest d-wave amplitude attained, although the increase in amplitude for flash durations greater than 75 ms was not statistically confirmed [18]. Further increase of the d-wave amplitude was observed with flashes up to 900 ms duration [19], but responses to such long durations take longer to record. Patient comfort is a consideration when selecting the light duration, also to minimize possible eye closure and blink artefacts.

b) Stimulus wavelength. The flash and background wavelength for the ISCEV-standard ERG are defined as visibly white, with CIE co-ordinates near x = 0.31, y = 0.32. Both white flashes and chromatic (blue and green) flashes have been used to elicit long-duration ERGs of similar waveform [18]. Some laboratories use orange stimuli in the presence of green background, for more selective stimulation of L-and M-cone system with simultaneous suppression of the rods and S-cones [17, 20]. These stimuli are effective at eliciting On- and Off- responses and have been shown to be informative in numerous studies (Table 1). Longer wavelength stimuli (red) may decrease the d-wave and change the shape of the b-wave [18, 19] and should be avoided.

c) Stimulus strength and background luminance. Brighter backgrounds require stronger stimuli to elicit detectable responses [18]. If stimuli are too weak responses are small. If stimuli are too strong the b-wave becomes broader and peak time variable and difficult to determine [18], while the d-wave becomes either decreased or dominated by a component of longer peak time, (the basis of the photopic hill phenomenon) [21]. Strong stimuli and backgrounds may also be poorly tolerated by some patients.

### 6. Calibration:

The protocol is technically similar to that for the ISCEV standard ERG, and the calibration and frequency of calibration should follow the latest ISCEV standard [1]. The strength of the

stimulus and background luminance should be specified in photopic candelas per meter squared (phot cd.m<sup>-2</sup>).

# 7. Protocol Specifications:

Patient preparation follows that for the current ISCEV standard ERG [1]. It is suggested that for routine applications the long-duration ERG is added to the ISCEV-standard protocol after the other LA ERGs. The following additional specifications are suggested:

a) Stimulus duration. It is suggested to use durations of 150ms or 200ms, to allow clear separation of On and Off responses, efficient signal averaging and for consistency with the majority of published clinical studies to date (Table 1).

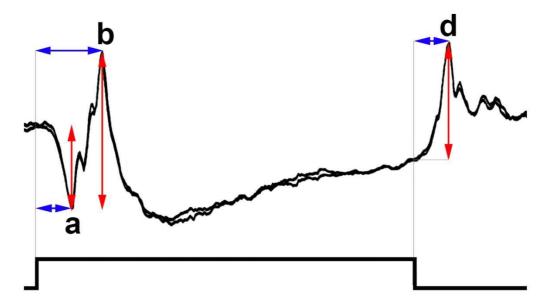
b) Stimulus wavelength. A white stimulus on a white background (as used in majority of published studies, Table 1) or a chromatic light on a chromatic background (e.g. to saturate S-cones) may be used providing longer wavelength (red) stimuli are avoided. Orange (620 nm) stimuli in the presence of green (560 nm) background may allow more selective stimulation of L-and M-cone systems with simultaneous suppression of the rods and S-cones.

c) Stimulus strength and background luminance. It is suggested that stimulus luminance for white stimuli is 250 cd.m<sup>-2</sup> or within the range phot 150-350 cd.m<sup>-2</sup> with a background luminance of 30 cd.m<sup>-2</sup>.

d) Inter-stimulus time (rate). A maximum rate of 0.5 per s ( $\leq$ 2.0 Hz) conforms to the current ISCEV Standard for the LA 3.0 ERG.

# 8. Response evaluation:

The long-duration flash ERG response consists of three prominent waves, the a-, and b-waves as part of the On- response and the Off- response complex, mainly the d-wave (Figure 1). A negative a-wave appears in response to the onset of the stimulus, followed by the positive-going On-response b-wave. The d-wave of the Off- response is the positive peak, which appears after the termination of the stimulus. The amplitude of the a-wave is measured from the baseline to the first negative trough. The amplitude of the b-wave is measured from the trough of the a-wave to the peak of the b-wave. The amplitude of the d-wave is measured from the time point of stimulus offset to the peak of the d-wave. The peak of each of the waves. The peak time of the d-wave is measured from the stimulus offset to the peak of the d-wave.



**Figure 1:** Diagram of the three main components of the long-duration flash ERG and their measurement. Stimulus duration is indicated at the bottom of the figure; red arrows indicate amplitude measurement, blue arrows indicate peak time measurement.

The amplitude and peak time values should be evaluated according to the reference values, which should be established in each laboratory for its own equipment, recording protocols and patient population.

## 9. Reporting:

This protocol is intended to be used for routine applications as an extension to the standard ERG protocol, and reporting should follow the latest ISCEV Standard for ERG [1]. Additionally the spectral characteristics of the stimulus and background should be acknowledged if different from the standard LA ERG (e.g. peak wavelength and bandwidth). The duration (in ms if different from 150 or 200ms) and luminance (in phot cd.m<sup>-2</sup>) of stimulus and background should be stated. Unless already embedded within the ISCEV standard ERG protocol pupil size and duration of light adaption should be stated. The amplitude of the a-, b- and d-waves and respective time to peaks may be reported along with age-appropriate laboratory reference normative data. It is acknowledged that in diagnostic studies involving ISCEV standard ERGs it may be sufficiently informative to describe the relative involvement of a-, b-, and d- waves qualitatively.

#### 10. Experimental procedures excluded from this guideline:

ERG recordings with long-duration flashes can be affected by technical issues, and blink and squint artefacts may disturb the recordings. Some authors suggest that this may be alleviated by using stimuli with sawtooth luminance profiles. The recordings are typically performed at photopic luminances. Instead of a flash upon a background, these stimuli are modulated around a mean. The stimulus strength is quantified by Michelson contrast (C):  $C = (L_{max} + L_{min})/(L_{max} - L_{min})$  in which  $L_{max}$  and  $L_{min}$  are the maximal and minimal luminance in the stimulus. Since this stimulus is given repetitively with a frequency between 2 and 8 Hz, blink artefacts do not play a large role. Higher frequencies are not recommended because the responses to subsequent stimuli may merge. On- and Off-responses are obtained separately

- by using rapid-ON and rapid-OFF stimuli. They have been used in a variety of disorders [22-
- 178 27] and may have benefits, but have not been widely available on commercial systems.
- On- and Off- responses might not be the same for all stimulus types. It has been reported
- that On- and Off- responses originating in the L-cones have the same morphology as those
- obtained with luminance stimuli. In contrast, M-cone driven On-responses resemble Off-
- responses with L-cone isolating and luminance stimuli and vice versa, suggesting that cone
- opponent processes may be involved [28-30].
- Beside the sawtooth stimulation, increment and decrement stimulation is also one of
- alternative ways for eliciting the d-wave without a major impact of blinking artefacts [31].
- 186 An alternative method extracts On- and Off- responses from the LA 3.0 ERG through the
- quantification of wavelet coefficients by discrete wavelet transform (DWT) analyses [32].
- This approach suggests the activity of the retinal On- pathway to be related to a 20Hz
- component of the photopic ERG, while the Off- pathway activity is reflected by a 40Hz
- component. This finding was based on the fact that 20Hz and 40Hz components of the
- photopic b-wave are selectively attenuated in case of imbalanced dysfunction of the On- and
- 192 Off- pathways in some diseases [32], confirmed with the DWT of photopic long-duration
- 193 flash ERGs [33].
- 194 Since long-duration flash ERG is potentially valuable ERG method in animal studies, the
- researchers should be aware that positive On- and Off- responses, as those in humans, can
- $\,\,$  196  $\,\,$  only be recorded in some non-human primates, while On- and Off- responses with
- electronegative waveform are presented in rodents including mice and rats [34].

# 199 **11. Relevant References:**

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- 1. McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R, Bach M (2015)
- 202 ISCEV Standard for full-field clinical electroretinography (2015 update). Doc Ophthalmol
- 203 130:1-12

204

- 205 **2.** Miyake Y, Yagasaki K, Horiguchi M, Kawase Y (1987) On- and off responses in photopic
- 206 electroretinogram in complete and incomplete types of congenital stationary night
- 207 blindness. Jpn J Ophthalmol 31:81-87

208

- 3. Alexander KR, Fishman GA, Peachey NS, Marchese AL, Tso MOM (1992) 'On' response
- defect in paraneoplastic night blindness with cutaneous malignant melanoma. Invest
- 211 Ophthalmol Vis Sci 33:477-483

212

- **4.** Sieving PA (1993) Photopic ON- and OFF-pathway abnormalities in retinal dystrophies.
- Trans Am Ophthalmol Soc 91:701-773

215

- 216 **5.** Bush RA, Sieving PA (1994) A proximal retinal component in the primate photopic ERG a-
- wave. Invest Ophthalmol Vis Sci 35:635-645

218

- 219 **6.** Sieving PA, Murayama K, Naarendorp F (1994). Push-pull model of the primate photopic
- 220 electroretinogram: a role for hyperpolarizing neurons in shaping the b-wave. Vis Neurosci
- 221 11:519-532

7. Ueno S, Kondo M, Ueno M, Miyata K, Terasaki H, Miyake Y (2006) Contribution of retinal
 neurons to d-wave of primate photopic electroretinograms. Vision Res 46:658-664

- 8. Khan NW, Kondo M, Hiriyanna KT, Jamison JA, Bush RA, Sieving PA (2005) Primate retinal
  signaling pathways: Suppressing ON-pathway activity in monkey with glutamate analogues
  mimics human CSNB1-NYX genetic night blindness. J Neurophysiol 93:481-492
- 230 9. Ruether K, Kellner U (1998) Inner retinal function in hereditary retinal dystrophies. Acta
  231 Anat (Basel) 162:169-177
- 233 10. Quigley M, Roy MS, Barsoum-Homsy M, Chevrette L, Jacob JL, Milot J (1996) On- and off 234 responses in the photopic electroretinogram in complete-type congenital stationary night
  235 blindness. Doc Ophthalmol 92:159-165
- **11.** Kellner U, Bornfeld N, Foerster MH (1995) Severe course of cutaneous melanoma
  238 associated paraneoplastic retinopathy. Br J Ophthalmol 79:746-752
  239
- 12. Thompson DA, Lyons RJ, Liasis A, Russell-Eggitt I, Jägle H, Grünewald S (2012) Retinal on pathway deficit in congenital disorder of glycosylation due to phosphomannomutase
  deficiency. Arch Ophthalmol 130:712-719
- **13.** Cibis GW, Fitzgerald KM (2001) The negative ERG is not synonymous with nightblindness.
  245 Trans Am Ophthalmol Soc 99:171-176
- 14. Allen LE, Zito I, Bradshaw K, Patel RJ, Bird AC, Fitzke F, Yates JR, Trump D, Hardcastle AJ,
  Moore AT (2003) Genotype-phenotype correlation in British families with X linked congenital
  stationary night blindness. Br J Ophthalmol 87:1413-1420
- **15.** Robson AG, Richardson EC, Koh AH, Pavesio CE, Hykin PG, Calcagni A, Graham EM,
  252 Holder GE (2005) Unilateral electronegative ERG of non-vascular aetiology. Br J Ophthalmol
  253 89:1620-1626
  - **16.** Kim JM, Payne JF, Yan J, Barnes CS (2012) Negative electroretinograms in the pediatric and adult population. Doc Ophthalmol 124:41-48
- **17.** Audo I, Robson AG, Holder GE, Moore AT (2008) The negative ERG: clinical phenotypes and disease mechanisms of inner retinal dysfunction. Surv Ophthalmol 53:16-40
  - **18.** Sustar M, Hawlina M, Brecelj J (2006) ON- and OFF-response of the photopic electroretinogram in relation to stimulus characteristics. Doc Ophthalmol 113:43-52
  - **19.** Evers HU, Gouras P (1986) Three cone mechanisms in the primate electroretinogram: two with, one without off-center bipolar responses. Vision Res 26:245-254
  - **20.** Audo I, Michaelides M, Robson AG, Hawlina M, Vaclavik V, Sandbach JM, Neveu MM, Hogg CR, Hunt DM, Moore AT, Bird AC, Webster AR, Holder GE (2008) Phenotypic variation in enhanced S-cone syndrome. Invest Ophthalmol Vis Sci 49:2082-2093

**21.** Kondo M, Piao CH, Tanikawa A, Horiguchi M, Terasaki H, Miyake Y (2000) Amplitude decrease of photopic ERG b-wave at higher stimulus intensities in humans. Jpn J Ophthalmol 44: 20-28

22. Barnes CS, Alexander KR, Fishman GA (2002) A distinctive form of congenital stationary night blindness with cone ON-pathway dysfunction. Ophthalmology 109:575-583

23. Dryja TP, McGee TL, Berson EL, Fishman GA, Sandberg MA, Alexander KR, Derlacki DJ,
 Rajagopalan AS (2005) Night blindness and abnormal cone electroretinogram ON responses
 in patients with mutations in the GRM6 gene encoding mGluR6. Proc Natl Acad Sci USA
 102:4884-4889

**24.** Alexander KR, Barnes CS, Fishman GA, Milam AH (2002) Nature of the cone ON-pathway dysfunction in melanoma-associated retinopathy. Invest Ophthalmol Vis Sci 43:1189-1197

**25.** Alexander KR, Fishman GA, Barnes CS, Grover S (2001) ON-Response deficit in the electroretinogram of the cone system in X-linked retinoschisis. Invest Ophthalmol Vis Sci 42:453-459

**26.** Pangeni Lämmer GR, Tornow RP, Horn FK, Kremers J (2012) On- and Off- response ERGs elicited by sawtooth stimuli in normal subjects and glaucoma patients. Doc Ophthalmol 124: 237-248.

**27.** Barboni MTS, Nagy BV, de Araújo Moura AL, Damico FM, da Costa MF, Kremers J, Ventura DF (2013) ON and OFF electroretinogram and contrast sensitivity in Duchenne muscular dystrophy. Invest Ophthalmol Vis Sci 54:3195-3204

**28.** McKeefry D, Kremers J, Kommanapalli D, Challa NK, Murray IJ, Maguire J, Parry NRA (2014) Incremental and decremental L- and M-cone driven ERG responses: I. Square-wave pulse stimulation. J Opt Soc Am A 31: A159-A169

**29.** Kremers J, Pangeni G, Tsaousis KT, McKeefry D, Murray IJ, Parry NRA (2014) Incremental and decremental L- and M-cone driven ERG responses: II. Sawtooth stimulation. J Opt Soc Am A 31: A170-A178

**30.** Tsai TI, Jacob MM, McKeefry D, Murray IJ, Parry NRA, Kremers J (2016) Spatial properties of L- and M-cone driven incremental (On-) and decremental (Off-) electroretinograms: evidence for the involvement of multiple post-receptoral mechanisms. J Opt Soc Am A 33: A1-A11

**31.** Vukmanic E, Godwin K, Shi P, Hughes A, DeMarco P Jr. Full-field electroretinogram response to increment and decrement stimuli (2014) Doc Ophthalmol 129:85-95

**32.** Gauvin M, Little JM, Lina JM, Lachapelle P (2015) Functional decomposition of the human ERG based on the discrete wavelet transform. J Vis 15:14

318 33. Gauvin M, Sustar M, Little JM, Brecelj J, Lina JM, Lachapelle P (2017) Quantifying the ON
 319 and OFF contributions to the flash ERG with the discrete wavelet transform. Transl Vis Sci
 320 Technol 10;6:3

**34.** Lei B (2003) The ERG of guinea pig (Cavis porcellus): comparison with I-type monkey

323 and E-type rat. Doc Ophthalmol 106:243-249

324

325 35. Kabanarou SA, Holder GE, Bird AC, Webster AR, Stanga PE, Vickers S, Harney BA (2004) 326 Congenital stationary night blindness and a "Schubert-Bornschein" type electrophysiology in 327 a family with dominant inheritance. Br J Ophthalmol 88:1018-1022

328

329 36. Constable PA, Gaigg SB, Bowler DM, Jägle H, Thompson DA (2016) Full-field 330 electroretinogram in autism spectrum disorder. Doc Ophthalmol 132:83-99

331

332 **37.** Sustar M, Perovšek D, Cima I, Stirn-Kranjc B, Hawlina M, Brecelj J (2015) 333 Electroretinography and optical coherence tomography reveal abnormal post-334 photoreceptoral activity and altered retinal lamination in patients with enhanced S-cone 335 syndrome. Doc Ophthalmol 130:165-177

336

337 38. Ba-Abbad R, Robson AG, Yap YC, Moore AT, Webster AR, Holder GE (2014) PRPH2 338 mutations as a cause of electronegative ERG. Retina 34:1235-1243

339

340 **39.** Schatz A, Breithaupt M, Hudemann J, Niess A, Messias A, Zrenner E, Bartz-Schmidt KU, 341 Gekeler F, Willmann G (2014) Electroretinographic assessment of retinal function during 342 acute exposure to normobaric hypoxia. Graefes Arch Clin Exp Ophthalmol 252:43-50

343 344

40. Raghuram A, Hansen RM, Moskowitz A, Fulton AB (2013) Photoreceptor and postreceptor responses in congenital stationary night blindness. Invest Ophthalmol Vis Sci 346 54:4648-4658

347

345

348 41. Vincent A, Robson AG, Neveu MM, Wright GA, Moore AT, Webster AR, Holder GE (2013) 349 A phenotype-genotype correlation study of X-linked retinoschisis. Ophthalmology 120:1454-350 1464

351 352

42. Moskowitz A, Hansen RM, Eklund SE, Fulton AB (2012) Electroretinographic (ERG) responses in pediatric patients using vigabatrin. Doc Ophthalmol 124:197-209

353 354 355

356

357

43. Kondo M, Sanuki R, Ueno S, Nishizawa Y, Hashimoto N, Ohguro H, Yamamoto S, Machida S, Terasaki H, Adamus G, Furukawa T (2011) Identification of autoantibodies against TRPM1 in patients with paraneoplastic retinopathy associated with ON bipolar cell dysfunction. PLoS One 6(5):e19911

358 359 360

361

44. Hankins MW, Jones SR, Jenkins A, Morland AB (2001) Diurnal daylight phase affects the temporal properties of both the b-wave and d-wave of the human electroretinogram. Brain Res 889:339-343

362 363 364

45. Shinoda K, Ohde H, Mashima Y, Inoue R, Ishida S, Inoue M, Kawashima S, Oguchi Y (2001) On- and off-responses of the photopic electroretinograms in X-linked juvenile retinoschisis. Am J Ophthalmol 131:489-494

366 367

365

368 46. Shinoda K, Ohde H, Ishida S, Inoue M, Oguchi Y, Mashima Y (2004) Novel 473-bp deletion 369 in XLRS1 gene in a Japanese family with X-linked juvenile retinoschisis. Graefes Arch Clin Exp 370 Ophthalmol 242:561-565

371

372 47. Yamamoto S, Hayashi M, Tsuruoka M, Ogata K, Tsukahara I, Yamamoto T, Takeuchi S 373 (2002) Selective reduction of S-cone response and on-response in the cone

electroretinograms of patients with X-linked retinoschisis. Graefes Arch Clin Exp Ophthalmol 240:457-460

48. Shinoda K, Ohde H, Inoue R, Ishida S, Mashima Y, Oguchi Y (2002) ON-pathwaydisturbance in two siblings. Acta Ophthalmol Scand 80:219-223

49. Koh AH, Hogg CR, Holder GE (2001) The incidence of negative ERG in clinical practice.
 Doc Ophthalmol 102:19-30

**50.** Khan NW, Jamison JA, Kemp JA, Sieving PA (2001) Analysis of photoreceptor function and inner retinal activity in juvenile X-linked retinoschisis. Vision Res 41:3931-3942

51. Imaizumi M, Matsumoto CS, Kimoto K, Furushima M, Nakatsuka K (2002) "On" responsedysfunction in multifocal posterior pigment epitheliopathy. Retina 22:33-36

52. Holder GE, Robson AG, Pavesio C, Graham EM (2005) Electrophysiological
 characterisation and monitoring in the management of birdshot chorioretinopathy. Br J
 Ophthalmol 89:709-718

53. Renner AB, Kellner U, Cropp E, Foerster MH (2006) Dysfunction of transmission in the
 inner retina: incidence and clinical causes of negative electroretinogram. Graefes Arch Clin
 Exp Ophthalmol 244:1467-1473

**54.** Hotta K, Kondo M, Nakamura M, Hotta J, Terasaki H, Miyake Y, Hida T. (2006) Negative electroretinograms in pericentral pigmentary retinal degeneration. Clin Exp Ophthalmol 34:89-92

**55.** Usui T, Tanimoto N, Ueki S, Miki A, Takagi M, Hasegawa S, Abe H (2005) Night blindness with depolarizing pattern of ON/OFF response in electroretinogram: a case report. Doc Ophthalmol 111:15-21

**56.** Sustar M, Stirn-Kranjc B, Hawlina M, Brecelj J (2008) Photopic ON- and OFF-responses in complete type of congenital stationary night blindness in relation to stimulus intensity. Doc Ophthalmol 117:37-46

**57.** Tanimoto N, Usui T, Ichibe M, Kuze M, Takagi M, Hasegawa S, Sato M, Tanaka K, Abe H (2006) Negative scotopic ERG and photopic ERG ON response impairment in a patient with normal dark adaptation. Doc Ophthalmol 113:171-177

58. Horn FK, Gottschalk K, Mardin CY, Pangeni G, Jünemann AG, Kremers J (2011) On and off
 responses of the photopic fullfield ERG in normal subjects and glaucoma patients. Doc
 Ophthalmol 122:53-62

**59.** Machida S, Ohguro H, Tateda M, Sato H, Kurosaka D (2011) Melanoma-associated retinopathy associated with intranasal melanoma. Doc Ophthalmol 122:191-197

# Part B. Justification for the protocol details and description of the consultation process

A literature review was performed with Medline search engine to find publications that reported the long-duration or On-Off ERG using following keywords: electroretinogram or full-field ERG or long duration flash, b-wave or ON-response, d-wave or OFF-response. Out of 110 matches, animal studies were excluded, as well as studies using multifocal, focal and sawtooth type of stimulation, as well as studies focusing on the function of rod or ganglion cell systems. Studies from the year 1986 to 2016 were reviewed and those with specified stimulus parameters are summarized in Table 1.

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Ref.	Stimulus strength	Background luminance	Stimulus duration	Stimulus and background wavelength
			background	
[3]	$3.7 \log cd/m^2$	$2.1 \log cd/m^2$	5-100 ms	white flash on white
	(5011 cd/m <sup>2</sup> )	(125 cd/m <sup>2</sup> )		background
[9, 53]	3 cd s/m <sup>2</sup>	10 cd/m <sup>2</sup>	200 and 250 ms	red and green flash on
	(15 cd/m <sup>2</sup> )			white background
[10]	750 cd/m <sup>2</sup>	42 cd/m <sup>2</sup>	500 ms	white flash on white
				background
[11]	3 cd s/m <sup>2</sup>	10 cd/m <sup>2</sup>	256 ms	630 nm flash on white
	$(12 \text{ cd/m}^2)$			background
[12]	200 cd/m <sup>2</sup>	43 cd/m <sup>2</sup>	90, 120ms	white 6500K
[13]	3 log cd s/m <sup>2</sup>	2 log cd/m <sup>2</sup>	200 ms	white flash on white
	$(5000 \text{ cd/m}^2)$	$(100 \text{ cd/m}^2)$		background
[14]	398 cd/m <sup>2</sup>	48 cd/m <sup>2</sup>	200 ms	white flash on white
				background
[21]	$0.6-3.5 \log cd/m^2$	40 cd/m <sup>2</sup>	250 ms	white flash on white
	(4-3162 cd/m <sup>2</sup> )			background
[18, 56]	$0.4 - 2.1 \log cd s/m^2$	20-50 cd/m <sup>2</sup>	5-200 ms	white, 460, 508 and 667
	(12.5-629 cd/m <sup>2</sup> )	,		nm flash on white
	, ,			background
15, 17, 20,	560 cd/m <sup>2</sup>	160 cd/m <sup>2</sup>	150-200 ms	620 nm flash on 530 nm
38, 41, 52]				background
[35]	440 cd/m <sup>2</sup>	160 cd/m <sup>2</sup>	200 ms	612 nm flash on 530 nm
[55]				background
[36]	133 cd/m <sup>2</sup>	43 cd/m <sup>2</sup>	120 ms	white 6500K
[37]	1.7 $\log \operatorname{cd} \operatorname{s/m}^2$	40 cd/m <sup>2</sup>	200 ms	white flash (6500 K) on
[3/]	(250 cd/m <sup>2</sup> )	10 00/111	200 1113	white background
[39]	1 cd s/m <sup>2</sup>	30 cd/m <sup>2</sup>	250 ms	white 6500K
	(4 cd/m <sup>2</sup> )	30 ca/111	250 1115	Winte 0500K
[40]	200 cd/m <sup>2</sup>	42 cd/m <sup>2</sup>	150 ms	white flash on white
	200 cu/111	42 ca/111	150 1115	background
[42]	4 log ph td	3.3 log ph td	150 ms	white flash on white
	(200 cd/m <sup>2</sup> )*	(40 cd/m <sup>2</sup> )*	130 1113	background
[43]	200 cd/m <sup>2</sup>	30 cd/m <sup>2</sup>	100 ms	white flash on white
	200 cu/111	30 cu/111	100 1113	background
[44]	225 cd/m <sup>2</sup>	30 cd/m <sup>2</sup>	188 ms	white flash on white
	225 CU/III	50 CU/111	100 1112	
[45 40]	200 od /22	40 ad /22	150	background
[45-48]	300 cd/m <sup>2</sup>	40 cd/m <sup>2</sup>	150 ms	white flash on white

				background
[49]	650 cd/m <sup>2</sup>	160 cd/m <sup>2</sup>	120, 200 ms	orange flash on green
				background
[50]	200 cd/m <sup>2</sup>	42 cd/m <sup>2</sup>	150 ms	white flash on white
				background
[51]	1700 cd/m <sup>2</sup>	28 cd/m <sup>2</sup>	125 ms	white flash on white
				background
[54]	360 cd/m <sup>2</sup>	40 cd/m <sup>2</sup>	100 ms	white flash on white
				background
[55, 57]	1120 cd/m <sup>2</sup>	$30 \text{ cd/m}^2$	200 ms	white flash on white
. , .	•	•		background
[58]	40,60,80 cd/m <sup>2</sup>	20 cd/m <sup>2</sup>	240 ms	white flash on white
	, , .	,		background
[59]	2.5 log cd/m <sup>2</sup>	40 cd/m <sup>2</sup>	150 ms	white flash on white
1	5 ,	•		background

 $<sup>43\</sup>overline{4}$  \* calculated for 8 mm pupil diameter